Clinical research

Randomized clinical trial evaluating the efficacy of synbiotic supplementation on serum endotoxin and trimethylamine N-oxide levels in patients with dyslipidaemia

Shekoufeh Salamat^{1,2}, Alireza Jahan-Mihan³, Mohammad Reza Tabandeh^{4,5}, Anahita Mansoori¹

¹Nutrition and Metabolic Diseases Research Centre, Clinical Sciences Research Centre, Ahvaz Jundishapur University of Medical Sciences, Ahvaz, Iran

²Student Research Committee, Ahvaz Jundishapur University of Medical Sciences, Ahvaz, Iran

³Department of Nutrition and Dietetics, University of North Florida, Jacksonville, FL, USA ⁴Department of Biochemistry and Molecular Biology, Faculty of Veterinary Medicine, Shahid Chamran University of Ahvaz, Ahvaz, Iran

⁵Stem Cells and Transgenic Technology Research Center, Shahid Chamran University of Ahvaz, Iran

Submitted: 19 December 2023; Accepted: 1 January 2024 Online publication: 1 February 2024

Arch Med Sci Atheroscler Dis 2024; 9: e18–e25 DOI: https://doi.org/10.5114/amsad/178106 Copyright © 2024 Termedia & Banach

Abstract

Introduction: Elevated serum endotoxin and trimethylamine N-oxide (TMAO) are associated with metabolic disorders including dyslipidaemia and insulin resistance. This study aimed to evaluate the impact of a 12-week treatment with a synbiotic supplement on serum endotoxin and TMAO levels in patients diagnosed with dyslipidaemia.

Material and methods: A total of 56 patients who met the study inclusion criteria were recruited in this randomized, double-blind clinical trial. Participants were randomly assigned into intervention and control groups and received either synbiotic or placebo sachets twice a day for 12 weeks. The sociodemographic data, food intake, physical activity, and anthropometric indices of participants were assessed before and after intervention. Serum endotoxin, TMAO, and fasting blood glucose (FBG) levels were measured at the baseline and end of the study.

Results: No significant difference in the baseline characteristics of participants in the 2 groups was observed. After the 12 weeks of intervention, the mean of serum endotoxin (p < 0.0001), TMAO (p < 0.0001), and FBG (p < 0.0001) was decreased in patients who received synbiotic supplements while no significant change was observed in the control group. Moreover, a significant positive correlation between changes in endotoxin (r = 0.41, p = 0.041) and TMAO (r = 0.40, p = 0.047) with FBG changes was observed. **Conclusions:** A significant reduction in serum endotoxin and TMAO levels, as well as improvements in FBG, following 12 weeks of supplementation with synbiotics, may offer a potential approach for improving metabolic status in patients with dyslipidaemia.

Key words: synbiotic, dyslipidaemia, endotoxin, trimethylamine n-oxide, gut microbiota, gut permeability.

Corresponding author:

Dr. Anahita Mansoori Nutrition and Metabolic Diseases Research Centre Clinical Sciences Research Centre Ahvaz Jundishapur University of Medical Sciences Ahvaz, Iran E-mail: mansoori-a@ajums. ac.ir

ATHEROSCLEROTIC DISEASES AMS

Randomized clinical trial evaluating the efficacy of synbiotic supplementation on serum endotoxin and trimethylamine N-oxide levels in patients with dyslipidaemia

Introduction

Dyslipidaemia, characterized by abnormal serum lipid profile, is a prevalent and significant risk factor for the development of cardiovascular diseases (CVDs) [1] and is closely associated with other metabolic disorders, such as insulin resistance [2]. The pathogenesis of dyslipidaemia is complex and multifactorial, involving genetic predisposition, dietary habits, sedentary lifestyles [3], and more recently, dysbiosis, and alterations in gut microbiota (GM) composition and its metabolites [4].

The GM, a complex ecosystem composed of trillions of microorganisms, including bacteria, viruses, fungi, and archaea [5], has emerged as a key player in various aspects of host physiology, including nutrient metabolism, immune function, and the maintenance of gut barrier integrity [6, 7]. The influence of the GM on host lipid metabolism is mediated through metabolites produced by the GM, including short-chain fatty acids (SCFA), secondary bile acids, trimethylamine (TMA), and lipopolysaccharide (LPS) or endotoxin [8].

In dysbiosis conditions, the intestinal barrier is disrupted, leading to increased gut permeability [9] and bacterial endotoxins can translocate into the systemic circulation, triggering inflammatory responses and metabolic dysfunction [10]. Increased serum endotoxin levels or low-grade endotoxaemia are biomarkers for the assessment of gut permeability [11] and have been observed in individuals who are at risk of atherosclerosis and CVDs [12].

TMAO, on the other hand, is a metabolite produced by the GM metabolism of dietary choline, betaine, and L-carnitine. These substances metabolize to TMA and are then transformed into TMAO by hepatic flavin monooxygenases (FMOs) [13]. Elevated TMAO can contribute to inflammation, impair vascular function and structure, inhibit reverse cholesterol transport, and induce dyslipidaemia [14].

Probiotics and prebiotics are suggested to improve dysbiosis, gut permeability, and subsequently, low-grade endotoxaemia [12], and also to decrease the serum TMAO [15] through the effect of GM on the epithelial barrier [9].

Synbiotics, a combination of probiotics and prebiotics, aim to restore a healthy GM balance and modulate the production of metabolites, potentially improving lipid metabolism and reducing cardiovascular risk [16].

To the best of our knowledge, this is the first study investigating the effects of synbiotic administration on serum levels of TMAO and LPS as indirect measures of gut permeability [11] novel metabolic biomarkers for CVD risk [17] in patients with dyslipidaemia. This study aimed to explore the potential effect of a 12-week synbiotic supplementation on TMAO, LPS, and FBG levels in patients diagnosed with dyslipidaemia while shedding light on the intricate connections between dysbiosis, metabolic disorders, and potential therapeutic strategies.

Material and methods

Ethics statements

This study was approved by the Ethics Committee at the Ahvaz Jundishapur University of Medical Sciences based on the ethical guidelines of the 1975 Declaration of Helsinki (Approval code: IR.AJUMS.REC.1400.581), and it was registered in the Iranian Registry of Clinical Trials (Registration reference: IRCT20180128038540N2).

All participants were informed about the study protocol, and a written consent form was signed by participants during their registration.

Study design and participants

This study was a randomized, double-blind, placebo-controlled clinical trial with a parallel design. The participants were selected from a pool of patients diagnosed with dyslipidaemia, who were referred to the nutrition clinic in Mahshahr, Iran, and met the following inclusion criteria: 1. Willingness to participate in the study; 2. Diagnosed with dyslipidaemia [18]; and 3. Adult individuals up to 60 years old. The patients were excluded from the study if any of the following criteria were detected at any time of the study procedure: 1. Frequent travel; 2. Using chemical or herbal lipid-lowering drugs; 3. Having familial dyslipidaemia; 4. Suffering from cardiovascular, kidney, liver, endocrine, gastrointestinal, malignancies, and autoimmune diseases; 5. Any significant changes in body weight and lifestyle in the last 6 months; and 6. Using tobacco, drugs, or alcohol.

The participant's sociodemographic status, including age, gender, ethnicity, marital status, educational level, occupational status, smoking, and drinking alcohol, were evaluated through a self-report sociodemographic questionnaire at the time of recruitment.

Food intake and physical activity assessment

To evaluate the food intake of the participants, a 3-day (non-consecutive) food record was used at the beginning and end of the study. Energy and nutrient intakes including total calories, carbohydrates, protein, fat, saturated fatty acids (SFA), and fibre were analysed using Nutritionist IV software (First Data Bank; Hearst Corp, San Bruno, CA, USA). Physical activity (PA) was assessed using a selfreported PA questionnaire [19] and was determined as metabolic equivalents of task hour/ day (METs-h/day). To calculate each participant's MET, times spent on each activity (h/day) were multiplied by their typical energy expenditures, expressed in terms of METs. Finally, these values were added together to give the MET-h/day score for each participant.

Anthropometric measurements

Anthropometric measurements, including height, weight, waist circumference (WC), and body fat percentage (BFP) were measured, and body mass index (BMI) was calculated at the beginning and the end of the study. Height was measured to the nearest 0.1 cm using a digital stadiometer (InBody, South Korea). Weight and BF were assessed in light clothes and without shoes by an electrical body composition analyser (InBody 270, South Korea). A measuring tape was used to measure WC to the nearest 0.1 cm at the midpoint between the lowest rib and the iliac crest.

Biochemical assessments

For biochemical assessments, blood samples (5 ml) after 10-12 h of overnight fasting at the beginning and after 12 weeks of intervention were taken. The serum was separated by centrifugation (3000 RPM for 15 min at 25°C) and kept refrigerated at -70° C for further analysis.

The Enzyme-Linked Immunosorbent Assay (ELISA) kits were applied for the determination of serum Endotoxin (MBS260730, MyBioSource, China) and serum levels of TMAO (MBS7269386, MyBioSource, China).

The serum level of fasting blood glucose (FBG) was assessed using a commercial test kit (Glucose-SL test kit, Henry Schein, Inc., USA) with an enzymatic colorimetric method.

Sample size

To estimate the necessary sample size, we used the equation for parallel interventional studies with an α error of 0.05 and a β error of 20% (power = 80%). Based on a previous study [20] and the parameter of total cholesterol, the sample size was determined to be n = 50, with 25 participants per group. To account for a projected 10% dropout rate, a total of 56 patients with dyslipidaemia (n = 28/group) were recruited after the screening process.

Intervention

Participants were allocated into a treatment or a control group by simple randomization method

(computer-generated random numbers). Participants in the treatment group received one sachet of synbiotic powder, and those in the control group received a placebo sachet, dissolved in a cup of water, twice a day (30 min before lunch and dinner) for 12 weeks. Each synbiotic sachet contained a total dose of 3×10^{10} colony forming units (CFU) of 6 probiotic microorganism species, which were chosen based on previous studies, including Lactobacillus (L.) acidophilus [21], L. fermentum [22], L. plantarum [23], Bifidobacterium (B.) longum [24], B. lactis [25], and Saccharomyces (S.) boulardii [26] plus 5 g of prebiotics including inulin and fructooligosaccharide (FOS) [27] manufactured by Faradaru Pharmaceutical Company (Tehran, Iran). Placebo sachets (5 g corn starch) were identical in shape, size, colour, and packaging to the synbiotic sachets. Patients and researchers were blinded to the interventions. To blind the individuals who participated in the intervention, the packages were coded by a person who was not involved in the study.

Participants were asked to keep their usual diet and PA during the study. The daily reminder messages were sent to participants to ensure that they did not forget the consumption of supplements. At the end of the study, the percentage of unconsumed sachets was determined, and the patients who consumed less than 90% of their sachets were excluded from the study.

Statistical analysis

The normality of data distribution was determined by the Kolmogorov-Smirnov test. To compare parametric variables between 2 groups the independent sample t-test was used, and to compare variables within the group before and after the intervention a paired sample t-test was applied. To control for confounder variables, an analysis of covariance (ANCOVA) was used. Pearson's coefficient correlation analysis was used to determine the correlation between changes in FBG with endotoxin and TMAO changes. All analysis was carried out by SPSS software version 28 (SPSS Inc., Chicago, Illinois, USA). Data were analysed using both intention-to-treat (ITT) and per protocol (PP) approach analysis. $P \le 0.05$ was considered statistically significant.

Results

Of 56 patients selected for the intervention (n = 28) and control groups (n = 28), 6 patients (3 patients of each group) were excluded from the study due to low compliance rates. A high compliance rate (90.0%) was seen in 50 patients who completed 12 weeks of study. The mean age of participants was 42.4 years (43.2 ±7.2 against

41.7 ±4.9 years, in the intervention and placebo groups, respectively, p = 0.4). All the participants were male because of logistic limitations.

No statistically significant differences were observed in anthropometric indices (weight, BMI, WC, BFP), nutrient intake (total calorie, carbohydrate, protein, fat, SFA, and fibre intakes), and PAL before and after the study between and within groups in the ITT and PP approaches (Table I).

As shown in Table II, in the intervention group, the mean of serum endotoxin and TMAO significantly decreased after 12 weeks of supplementation. However, no significant differences were observed in the control group. Moreover, at the end of the study, a significant reduction was observed in FBG levels in the synbiotic group. Interestingly, Pearson's coefficient correlation analysis showed a significant positive correlation between FBG changes with endotoxin and TMAO changes (Table III).

In this study, we did not observe any side effects of synbiotic supplementation in either group.

Discussion

To the best of our knowledge, this is the first study to evaluate the effect of synbiotic supplementation on serum levels of TMAO and endotoxin in patients with dyslipidaemia. In this randomized controlled trial, administration of a multi-species synbiotic supplement (containing a total dose of 3×10^{10} CFU probiotic microorganisms including 5 bacteria and one yeast species, and 5 g prebiotics) in patients with dyslipidaemia

Variables		Intervention $(n = 25)$	Placebo (n = 25)	P-value*
BMI [kg/m²]	Before	30. 1 ±4.9	28.7 ±4.1	0.30
	After	30.2 ±4.9	28.8 ±4.1	0.31
	P-value*	0.40	0.32	
WC [cm]	Before	104.3 ±10.7	101.7 ±8.8	0.36
	After	104.4 ±10.6	102.0 ±8.4	0.60
	P-value	0.80	0.47	
BF (%)	Before	32.9 ±6.5	31.1 ±5.6	0.29
	After	33.3 ±7.2	30.9 ±5.5	0.19
	P-value	0.27	0.56	
Calories intake [kcal]	Before	2974±482	2861±441	0.51
	After	3118.8±557	2980/3±453	0.44
	P-value	0.31	0.58	
CHO [gr]	Before	418.8 ±85.0	397.7±72.4	0.41
	After	433 ±89.2	411 ±81.2	0.46
	P-value	0.32	0.59	
PRO [gr]	Before	118.9 ±18.4	115.7±16.4	0.74
	After	125.5 ±19.7	117.7 ±17.2	0.20
	P-value	0.31	0.88	
Fat [gr]	Before	111.6 ±30.1	99.2±14.9	0.11
	After	114.7±32.2	105.4±18.8	0.29
	P-value	0.66	0.41	
SFA [gr]	Before	28.4 ±9.9	28.1±8.3	0.91
	After	29.5±10.8	29.0±8.2	0.87
	P-value	0.54	0.70	
Fiber [gr]	Before	21.3 ±4.6	20.7±4.6	0.51
	After	22.6±5.1	20.9±4.8	0.33
	P-value	0.21	0.93	
PAL [Met/h]	Before	1.47	1.40	0.25
	After	1.47	1.43	0.46
	P-value	0.78	0.12	

 Table I. Patients' anthropometric indices, nutrient intake, and PAL before and after the study

BMI – body mass index, WC – waist circumference, BF – body fat, CHO – carbohydrate, Pro – protein, SFA – saturated fatty acids, PAL – physical activity level. *P-value was reported based on an independent samples t-test. *P-value was reported based on paired sample t-test. Data are presented as mean ± standard deviation. P-value < 0.05 is statistically significant. Shekoufeh Salamat, Alireza Jahan-Mihan, Mohammad Reza Tabandeh, Anahita Mansoori

Variables		Intervention $(n = 25)$	Placebo ($n = 25$)	P-value*
Endotoxin [EU/ml]	Before	3.94 ±0.88	3.56 ±1.02	0.16
	After	3.05 ±0.79	3.64 ±0.83	0.012
	P-value*	< 0.0001	0.66	
TMAO [ng/ml]	Before	90.6±30.9	91.1 ±24.7	0.90
	After	76.7 ±22.4	87.5 ±26.5	0.12
	P-value	< 0.0001	0.11	
FBG [mg/dl]	Before	106.2 ±21.5	99.2 ±30.1	0.35
	After	93.5 ±17.0	97.3 ±24.8	0.53
	P-value	> 0.0001	0.26	

Table II. Serum endotoxin, TMA, and FBG levels before and after intervention

TMAO - trimethylamine N-oxide, FBG - fasting blood. 'P-value was reported based on an independent samples t-test. 'P-value was reported based on paired sample t-test. Data are presented as mean ± standard deviation. P value < 0.05 is statistically significant.

Table III. Pearson's correlation coefficients of changes in FBG with the changes in serum LPS and TMAO levels in the studied groups

Parameter	Interve	ention	Placebo	
	P-value	r	<i>P</i> -value	r
LPS	0.041	0.41	0.41	0.17
ΤΜΑΟ	0.047	0.40	0.12	0.32

TMAO – trimethylamine N-oxide, LPS – lipopolysaccharide. Statistical analysis was done using Pearson's coefficient correlation. P-value < 0.05 is statistically significant.

resulted in a significant decrease in serum endotoxin and TMAO in the intervention group. Moreover, our results showed a significant reduction in FBG levels in patients who received synbiotic supplements, and a significant positive correlation between FBG changes and serum endotoxin and TMAO changes.

Consistent with our results, Tenore et al. showed consumption of 125 g/day lacto-fermented Annurca apple puree for 16 weeks (each dose contained about 3 × 10⁸ CFU L. rhamnosus LRH11 and L. plantarum SGL07) by individuals with CVD risk factors resulted in a decrease in TMAO level [28]. Moreover, Matsumoto et al. reported that supplementation with probiotic B. animalis subsp. Lactis LKM512 in healthy subjects could reduce TMA levels [29]. The results of another study by Moludi et al. showed that probiotic supplementation (containing 1.6 × 10⁹ CFU of L. rhamnosus LGG) along with calorie restriction in patients suffering coronary artery disease for 12 weeks led to a significant decrease in IL-1 β and endotoxin levels [30]. The findings of a study on psoriasis patients showed that supplementation with multi-strain probiotic capsules of at least 1.6 × 10⁹ CFU/g of probiotics including *L. acidophi*lus, B. bifidum, B. lactis, and B. langum for 8 weeks results in a significant reduction in serum endotoxin, hs-CRP, and IL-1ß levels [31]. In contrast, other studies did not find any significant effect of probiotic/prebiotic supplementation on TMAO or endotoxin levels. Boutagy et al. reported that treatment with the multi-strain probiotic VSL#3

for 4 weeks in non-obese males did not influence plasma TMAO concentrations following a high-fat diet [32]. The finding of another study showed that supplementation with 3×10^9 CFU *L. casei shirota* for 3 months in patients with metabolic syndrome increased LPS-binding protein (LBP) levels [33]. Moreover, Tripolt *et al.* reported that treatment with 6.5 × 10⁹ *L. casei Shirota* 3 times a day for 12 weeks did not decrease plasma levels of TMAO in patients with metabolic syndrome [34]. The inconsistency seen in the results of these studies could be explained by various dosages and species of probiotic microbes, the duration of the studies, the characteristics of participants, and various clinical settings [30].

Previous studies have shown that supplementation with the appropriate probiotic species or combination of probiotics can lead to modulation of the GM that in turn promotes the production of metabolites that can inhibit the TMAO synthesis pathway [35, 36]. It has been suggested that probiotics can produce bacteriocins that suppress pathogenic bacteria and modulate the anti-apoptotic and proliferation responses of intestinal epithelial cells [37, 38]. Furthermore, probiotics can protect intestinal epithelial cells from oxidative stress by secreting proteins that induce cytoprotective heat shock proteins [39].

The improvement in FBG levels following synbiotic supplementation is consistent with the known association between gut dysbiosis and metabolic disorders [40]. Recent research has suggested the role of dysbiosis in developing and progressing dysRandomized clinical trial evaluating the efficacy of synbiotic supplementation on serum endotoxin and trimethylamine N-oxide levels in patients with dyslipidaemia

lipidaemia and related metabolic disorders [41]. Individuals with hyperlipidaemia showed an increase in serum levels of LPS and TMAO [42]. It is evident that synbiotics can lead to alterations in GM composition, favouring the growth of beneficial bacteria [43] and increasing the production of SCFAs [44]. SCFAs improve gut barrier function and reduce gut permeability, which can limit the entry of LPS into the bloodstream, thereby reducing low-grade inflammation and insulin resistance [45]. Furthermore, SCFAs can stimulate the release of incretin hormones, including glucagon-like peptide-1 (GLP-1) and glucose-dependent insulinotropic polypeptide (GIP). These hormones play a crucial role in regulating blood glucose levels by promoting insulin secretion from pancreatic beta cells in response to elevated glucose levels. GLP-1 and GIP also inhibit glucagon release from pancreatic α cells, further contributing to glucose control [46].

We also found a positive correlation between changes in FBG and changes in endotoxin and TMAO levels. Aligned with our results, the study reported a positive correlation between serum endotoxin and FBG, and a negative association with serum HDL-C in patients with type 2 diabetes [47]. Our findings further support the potential interplay between GM and metabolic parameters. Our previous study showed that synbiotic supplementation significantly increased the abundance of beneficial gut bacteria and serum concentration of HDL-C in patients with dyslipidaemia [48]. The reduction in endotoxin, TMAO, and FBG levels following synbiotic supplementation suggests that targeting GM through synbiotic interventions could be a promising approach for managing dyslipidaemia and related metabolic disorders including insulin resistance.

Our study has several limitations. Firstly, the sample size was determined based on cholesterol parameters due to the limited existing studies on TMA and LPS at the time of study design. Secondly, only male patients were recruited, which limits the generalizability of our findings to the broader population. Lastly, we did not measure inflammatory biomarkers.

However, this study has some strengths. Firstly, it employed a rigorous randomized controlled trial design and was conducted on a homogeneous group of non-smoking men with similar lifestyles and ethnicities. Secondly, a high compliance rate was observed in participants who completed the 12-week study in both groups. Lastly, our study is the first to evaluate the effect of synbiotic supplementation on TMAO and LPS levels as novel biomarkers of CVD in patients diagnosed with dyslipidaemia.

In conclusion, the results of our study showed that a 12-week synbiotic supplementation in patients with dyslipidaemia resulted in significant reductions in serum endotoxin and TMAO levels, as well as improvements in FBG. Synbiotics might be a potential strategy for the management of metabolic disorders and the prevention of CVDs in adult men with dyslipidaemia. Future studies with larger and more diverse populations and a comprehensive assessment would be beneficial to further investigate the effects of synbiotic supplementation on metabolic status in patients with dyslipidaemia.

Acknowledgments

This study is part of a Ph.D. thesis funded by the Vice-Chancellor for Research and Technology of Ahvaz Jundishapur University of Medical Sciences (Grant number: NRC-0009).

Conflict of interest

The authors declare no conflict of interest.

References

- 1. Salamat S, Sharif SS, Nazary-Vanani A, Kord-Varkaneh H, Clark CC, Mohammadshahi M. The effect of green coffee extract supplementation on serum oxidized LDL cholesterol and total antioxidant capacity in patients with dyslipidemia: a randomized, double-blind, placebo-controlled trial. Eur J Integrative Med 2019; 28: 109-13.
- 2. Hirano T. Pathophysiology of diabetic dyslipidemia. J Atheroscler Thromb 2018; 25: 771-82.
- 3. Mosca S, Araújo G, Costa V, et al. Dyslipidemia diagnosis and treatment: risk stratification in children and adolescents. J Nutrition Metabol 2022; 2022: 4782344.
- 4. Flaig B, Garza R, Singh B, Hamamah S, Covasa M. Treatment of dyslipidemia through targeted therapy of gut microbiota. Nutrients 2023; 15: 228.
- 5. Hou K, Wu ZX, Chen XY, et al. Microbiota in health and diseases. Signal Transduct Target Ther 2022; 7: 135.
- 6. Jandhyala SM, Talukdar R, Subramanyam C, Vuyyuru H, Sasikala M, Reddy DN. Role of the normal gut microbiota. World J Gastroenterol 2015; 21: 8787-803.
- 7. Valdes AM, Walter J, Segal E, Spector TD. Role of the gut microbiota in nutrition and health. BMJ 2018; 361.
- 8. Schoeler M, Caesar R. Dietary lipids, gut microbiota and lipid metabolism. Rev Endocr Metabol Disord 2019; 20: 461-72.
- 9. Moludi J, Maleki V, Jafari-Vayghyan H, Vaghef-Mehrabany E, Alizadeh M. Metabolic endotoxemia and cardiovascular disease: a systematic review about potential roles of prebiotics and probiotics. Clin Exp Pharmacol Physiol 2020; 47: 927-39.
- 10. Khan A, Ding Z, Ishaq M, et al. Understanding the effects of gut microbiota dysbiosis on nonalcoholic fatty liver disease and the possible probiotics role: recent updates. Int J Biol Sci 2021; 17: 818-33.
- 11. Sochaczewska D, Ziętek M, Dołęgowska B, Kordek A, Szczuko M. Implications of indirect biomarkers of intestinal permeability in the stools of newborns and infants with perinatal risk factors for intestinal colonization disorders and infant feeding patterns. Nutrients 2022; 14: 2224.
- 12. Violi F, Cammisotto V, Bartimoccia S, Pignatelli P, Carnevale R, Nocella C. Gut-derived low-grade endotoxaemia,

atherothrombosis and cardiovascular disease. Nat Rev Cardiol 2023; 20: 24-37.

- Yang S, Li X, Yang F, et al. Gut microbiota-dependent marker TMAO in promoting cardiovascular disease: inflammation mechanism, clinical prognostic, and potential as a therapeutic target. Front Pharmacol 2019; 10: 1360.
- Wang Z, Tang WW, Buffa JA, et al. Prognostic value of choline and betaine depends on intestinal microbiota-generated metabolite trimethylamine-N-oxide. Eur Heart J 2014; 35: 904-10.
- Din AU, Hassan A, Zhu Y, Yin T, Gregersen H, Wang G. Amelioration of TMAO through probiotics and its potential role in atherosclerosis. Appl Microbiol Biotechnol 2019; 103: 9217-28.
- Jiang H, Cai M, Shen B, Wang Q, Zhang T, Zhou X. Synbiotics and gut microbiota: new perspectives in the treatment of type 2 diabetes mellitus. Foods 2022; 11: 2438.
- Ufnal M, Pham K. The gut-blood barrier permeability a new marker in cardiovascular and metabolic diseases? Med Hypotheses 2017; 98: 35-7.
- Tabatabaei-Malazy O, Qorbani M, Samavat T, Sharifi F, Larijani B, Fakhrzadeh H. Prevalence of dyslipidemia in Iran: a systematic review and meta-analysis study. Int J Prev Med 2014; 5: 373.
- 19. Aadahl M, Jørgensen T. Validation of a new self-report instrument for measuring physical activity. Med Sci Sports Exerc 2003; 35: 1196-202.
- 20. Cicero AF, Fogacci F, Bove M, Giovannini M, Borghi C. Impact of a short-term synbiotic supplementation on metabolic syndrome and systemic inflammation in elderly patients: a randomized placebo-controlled clinical trial. Eur J Nutr 2021; 60: 655-63.
- 21. Wang L, Guo MJ, Gao Q, et al. The effects of probiotics on total cholesterol: a meta-analysis of randomized controlled trials. Medicine 2018; 97 e9679.
- 22. Kullisaar T, Zilmer K, Salum T, Rehema A, Zilmer M. The use of probiotic L fermentum ME-3 containing Reg'Activ Cholesterol supplement for 4 weeks has a positive influence on blood lipoprotein profiles and inflammatory cytokines: an open-label preliminary study. Nutr J 2016; 15: 93.
- Fuentes MC, Lajo T, Carrión JM, Cuné J. Cholesterol-lowering efficacy of Lactobacillus plantarum CECT 7527, 7528 and 7529 in hypercholesterolaemic adults. Br J Nutr 2013; 109: 1866-72.
- 24. Xiao J, Kondo S, Takahashi N, et al. Effects of milk products fermented by Bifidobacterium longum on blood lipids in rats and healthy adult male volunteers. J Dairy Sci 2003; 86: 2452-61.
- 25. Bernini LJ, Simão ANC, Alfieri DF, et al. Beneficial effects of Bifidobacterium lactis on lipid profile and cytokines in patients with metabolic syndrome: a randomized trial. Effects of probiotics on metabolic syndrome. Nutrition 2016; 32: 716-9.
- 26. Gadelha C, Bezerra AN. Effects of probiotics on the lipid profile: systematic review. J Vasc Bras 2019; 18: e20180124.
- 27. Hadi A, Ghaedi E, Khalesi S, Pourmasoumi M, Arab A. Effects of synbiotic consumption on lipid profile: a systematic review and meta-analysis of randomized controlled clinical trials. Eur J Nutr 2020; 59: 2857-74.
- 28. Tenore GC, Caruso D, Buonomo G, et al. Lactofermented annurca apple puree as a functional food indicated for the control of plasma lipid and oxidative amine levels: results from a randomised clinical trial. Nutrients 2019; 11: 122.

- 29. Matsumoto M, Kitada Y, Shimomura Y, Naito Y. Bifidobacterium animalis subsp. lactis LKM512 reduces levels of intestinal trimethylamine produced by intestinal microbiota in healthy volunteers: a double-blind, placebo-controlled study. J Funct Foods 2017; 36: 94-101.
- 30. Moludi J, Kafil HS, Qaisar SA, Gholizadeh P, Alizadeh M, Vayghyan HJ. Effect of probiotic supplementation along with calorie restriction on metabolic endotoxemia, and inflammation markers in coronary artery disease patients: a double blind placebo controlled randomized clinical trial. Nutr J 2021; 20: 47.
- 31. Moludi J, Khedmatgozar H, Tabrizi FPF, Razmi H, Amirpour M, Fathi P. Skin-gut axis in psoriasis setting and plan for action: the effect of probiotics supplementation on clinical outcomes, metabolic endotoxemia, inflammation, and cardiovascular risk in patients with psoriasis. J Drugs Dermatol 2022; 21: 637-44.
- Boutagy NE, Neilson AP, Osterberg KL, et al. Probiotic supplementation and trimethylamine-N-oxide production following a high-fat diet. Obesity 2015; 23: 2357-63.
- 33. Leber B, Tripolt N, Blattl D, et al. The influence of probiotic supplementation on gut permeability in patients with metabolic syndrome: an open label, randomized pilot study. Eur J Clin Nutr 2012; 66: 1110-5.
- 34. Tripolt NJ, Leber B, Triebl A, Köfeler H, Stadlbauer V, Sourij H. Effect of Lactobacillus casei Shirota supplementation on trimethylamine-N-oxide levels in patients with metabolic syndrome: an open-label, randomized study. Atherosclerosis 2015; 242: 141-4.
- 35. Nagpal R, Wang S, Ahmadi S, et al. Human-origin probiotic cocktail increases short-chain fatty acid production via modulation of mice and human gut microbiome. Sci Rep 2018; 8: 12649.
- Skelly AN, Sato Y, Kearney S, Honda K. Mining the microbiota for microbial and metabolite-based immunotherapies. Nat Rev Immunol 2019; 19: 305-23.
- 37. Tao Y, Drabik KA, Waypa TS, et al. Soluble factors from Lactobacillus GG activate MAPKs and induce cytoprotective heat shock proteins in intestinal epithelial cells. Am J Physiol Cell Physiol 2006; 290: C1018-30.
- Yan F, Cao H, Cover TL, Whitehead R, Washington MK, Polk DB. Soluble proteins produced by probiotic bacteria regulate intestinal epithelial cell survival and growth. Gastroenterology 2007; 132: 562-75.
- 39. Forsyth CB, Farhadi A, Jakate SM, Tang Y, Shaikh M, Keshavarzian A. Lactobacillus GG treatment ameliorates alcohol-induced intestinal oxidative stress, gut leakiness, and liver injury in a rat model of alcoholic steatohepatitis. Alcohol 2009; 43: 163-72.
- Carding S, Verbeke K, Vipond DT, Corfe BM, Owen LJ. Dysbiosis of the gut microbiota in disease. Microbial Ecology Health Dis 2015; 26: 26191.
- 41. Wang PX, Deng XR, Zhang CH, Yuan HJ. Gut microbiota and metabolic syndrome. Chin Med J 2020; 133: 808-16.
- 42. Jia X, Xu W, Zhang L, Li X, Wang R, Wu S. Impact of gut microbiota and microbiota-related metabolites on hyperlipidemia. Front Cell Infect Microbiol 2021; 11: 634780.
- 43. Wang X, Yang J, Qiu X, et al. Probiotics, pre-biotics and synbiotics in the treatment of pre-diabetes: a systematic review of randomized controlled trials. Front Public Health 2021; 9: 645035.
- 44. McLoughlin RF, Berthon BS, Jensen ME, Baines KJ, Wood LG. Short-chain fatty acids, prebiotics, synbiotics, and systemic inflammation: a systematic review and meta-analysis. Am J Clin Nutr 2017; 106: 930-45.

Randomized clinical trial evaluating the efficacy of synbiotic supplementation on serum endotoxin and trimethylamine N-oxide levels in patients with dyslipidaemia

- 45. Den Besten G, Van Eunen K, Groen AK, Venema K, Reijngoud DJ, Bakker BM. The role of short-chain fatty acids in the interplay between diet, gut microbiota, and host energy metabolism. J Lipid Res 2013; 54: 2325-40.
- 46. Canfora EE, Jocken JW, Blaak EE. Short-chain fatty acids in control of body weight and insulin sensitivity. Nat Rev Endocrinol 2015; 11: 577-91.
- 47. Al-Attas OS, Al-Daghri NM, Al-Rubeaan K, et al. Changes in endotoxin levels in T2DM subjects on anti-diabetic therapies. Cardiovasc Diabetol 2009; 8: 20.
- 48. Salamat S, Tabandeh MR, Jahan-Mihan A, Mansoori A. The effect of supplementation with a multi-species synbiotic on serum lipid profile, abundance of beneficial gut bacteria and firmicutes to bacteroidetes ratio in patients with dyslipidemia; a randomized, double-blind, placebo-controlled, clinical trial. PharmaNutrition 2023: 100367.